SAMPLING AND ANALYSIS

Environmental Sampling and Remediation

EVALUATING THE PROBLEM -ENVIRONMENTAL SAMPLING

- Characterize source
- Environmental variability
- Sampling
 - Bulk
 - Air
 - Tape

• There are many limitations to sampling

INITIAL INVESTIGATION

Site history and background

• Visual observation

- Indoor and outdoor
- sites of water and moisture
- observed mold growth
- "moldy odors"



- Visual inspection good tool for assessing indoor moisture and fungal growth
- Sampling may be indicated when results of visual inspection are ambiguous or detailed information is necessary

HUMAN HEALTH EFFECTS AND SAMPLING

- Dose response not well defined concentrations in environment, species and human health responses all vary
- No universal agreement or recommended limits for fungi in indoor air

MOISTURE TESTING

Use of moisture detectors

- probes
- penetrating sensors

Pro's and Con's

- Relative measurement of moisture content moisture activity - no indication of mold growth only that moisture is present
- Potential inaccuracies/false readings

THERMAL IMAGING

- An useful tool to detect anomalies in building components. Water holds heat longer and results in water-damaged building materials retaining heat longer than adjoining or similar dry substrate.
 - Time saving
 - Examination of difficult to access areas
 - Examination of large areas quickly

THERMAL IMAGING

- Disadvantages
 - False readings
 - Cold and warm air can create false positives and false negatives
 - Warm may mask wet materials by warming surface
 - Cold air can cause a thermal pattern indicative of moisture
 - Reflections can distort the thermal pattern
 - Locating moisture behind ceramic tiles and some floor and wall covering can be difficult

• Use in conjunction with a moisture meter

EXAMPLE OF AN IR IMAGE



SAMPLING PROTOCOLS

Sampling locations

- Problem and non problem areas
- Outdoor samples
- Data represents a moment in time

Observe Site Conditions

- Note weather, activity levels, furnishings, plants
- HVAC operation
- Accessibility to outside air doors, windows

DESTRUCTIVE TESTING IN WALL CAVITY

- Wall cavity is a potential reservoir for mold and moisture
- The presence of mold alone does not necessarily mean the occupied space is contaminated
- Opening wall cavities may create indoor contamination
- Opening wall cavity from the exterior

WALL CAVITIES

Inspections

- Snap shot of inside of wall only
- Molds on lumber surfaces is typical
- Spores from dusts, etc.
- Collecting air samples from inside
 - Often done, typically not useful
 - No comparison criteria (normal vs. abnormal)
 - May lead to false indication of water intrusion/fungal growth



SURFACE SAMPLING

- Tape, Swab or Bulk
- Direct microscopic exam
 - To determine and identify fungi on surface
 - Presence of mold spores
 - Mold growing or may have been growing
 - Indicator of a fungal reservoir
 - Marker genera indicative of indoor mold growth

SURFACE SAMPLING

• Disadvantages

- Not a direct indication of what might be airborne
- Sometimes results are interpreted wrong or poorly may vary depending where sample is taken (i.e., dusty area)
- Not all the spores seen under the microscope may be viable (alive).
- It may be advisable to combine direct exam samples with culture methods to get a better picture of what live molds are present.

SPORE TRAP MONITORING

Advantages

- Capture majority of spores/particulate matter in air
- Accurately characterize a problem
- Quantify pollen, skin cells, hyphal fragments, other particles

• <u>Disadvantages</u>

- Some spores difficult to identify Pen/Asp
- Viability not assessed not typically critical
- High variability
- Confounding issues high concentrations not necessarily indicative of a problem
- Differences in interpretation



SPORE TRAP REPORT: NON-VIABLE METHODOLOGY Instrument Used: Zefon Air-O-Cell volumetric air sampler

Location: MB061799 –	4: Portable Bldg. #28, afternoon, inside		5: Portable Bldg. #28, afternoon, inside		6: Portable Bldg. #28, afternoon, outside	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria	60	1,325	112	2,472	84	1,854
Arthrinium						
Ascospores*	100				Constant Solo	
Aureobasidium pullulans						
Basidiospores*			4	88	8	177
Botrytis						
Chaetomium						
Cladosporium	3,115	68,764	6,440	142,163	5,740	126,711
Curvularia				-		
Drechslera/Bipolaris group					1	22
Epicoccum			1	22		
Fusarium						
Myrothecium						
Nigrospora						
Other colorless						
Penicillium/Aspergillus types†	4	88			40	883
Pithomyces						•
Rusts*			1	22	2	44
Smuts*, Periconia, Myxomycetes*	6	132	20	442	18	397
Stachybotrys						
Stemphylium	1	22	2	44	2	44
Torula herbarum			1	22		
Ulocladium						
Unknown						
Zygomycetes (possible)					tening and a second	
Background debris (1-4+) ^{††}	3+		. 4		3	
Sample volume (liters)	45.3		45.3		45.3	
TOTAL SPORES/M3		70,331		145,275		130,132

Comments:

CULTURABLE AIR SAMPLING

- Air drawn through an impactor plate with a petri dish below
- Spores incubate and grow on agar
- Viable fungi
- Indoor vs. outdoor air comparison
- Will differentiate species Pen/Asp
- May be used for bacterial count

CULTURABLE AIR SAMPLING

- Does not indicate non-viable spores/fragments
- 5-7 days for incubation
- Short sampling period (minutes)
- Fungi and bacteria present in the air may not be as well represented in culture - competition
- Some microbes do not grow well or at all on the culture media.

CULTURABLE MOLD SPORE REPORT Instrument Used: Andersen volumetric air sampler

Location: MB061799 –	5: Portable Bldg. #28, afternoon, inside		6: Portable Bldg. #28, afternoon, outside		BL: Blank	
	raw ct.	cfu/m3	raw ct.	cfu/m3	raw ct.	cfu/m3
Acremonium						
Alternaria	2	35	5	88		
Aspergillus, other						
Aspergillus ochraceous						
Aspergillus flavus					and the second	
Aspergillus fumigatus						
Aspergillus nidulans						
Aspergillus niger						
Aspergillus versicolor						
Aureobasidium						
Beauveria						
Botrytis						
Chaetomium						
Cladosporium	195	13.039	200	> 20.777		
Curvularia				20,777		
Drechslera						
Epicoccum	1	18	1	18		
Fusarium			-	10		
Mucor / Rhizopus			1	18		
Paecilomyces			-	10		-
Penicillium	1	18	16	300		
Phoma						
Sporobolomyces						
Stachybotrys						
Trichoderma						
Ulocladium						
Nonsporulating colonies	8	141	15	283		
Sample volume (liters)	56.6		56.6	200		No
FOTAL CFU*/M3		13,251	50.0	> 21 484		growth

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DATA INTERPRETATION

- Compare complaint and control samples
- Relative quantities
- Rank order of prevalence
- Unusual, allergenic types

POLYMERASE CHAIN REACTION SEQUENCE DETECTION (PCR)

Advantages

- Developed by EPA uses fungal DNA
- Quick turnaround 1 to 2 days
- Accurate fungal detection (whether viable or not)
- Will allow fungal differentiation where spore trap may not
- Good detection sensitivity
- Longer sampling period



Disadvantages

- Expensive compared to spore trap
- Not best for screening purposes as is spore trap or culturable (only detects fungi requested)
- Cannot distinguish viable from non-viable
- Currently not a large data base of indoor fungi
- No standards or guidelines still must rely on indoor vs. outdoor results



• Often done by "mold inspectors"

- Little basis in scientific validity
- Data cannot be interpreted
- Provides little information regarding conditions or fungal reservoirs
- Sampling is conducted with no or incomplete objectives regarding what to do with the data

WHEN AND WHY TO SAMPLE

• Only if it's done for a good reason

- Demonstrate exposure when a health effect has already been established
- Link to unknown reservoir
- Post-remediation clearance sampling
- Know what you want from the data (hypothesis) before you design your sampling strategy
- Respect the limitations of the data
- Sampling is at times overdone with little forethought paid to interpreting the data